

Observing Single Molecules Complexing with Cucurbit[7]uril through Nanogap Surface-Enhanced Raman Spectroscopy

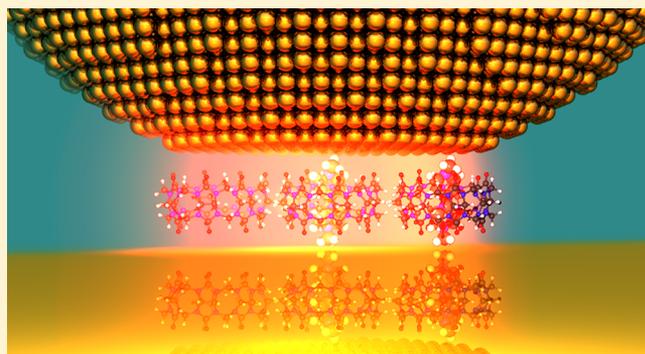
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S Supporting Information

ABSTRACT: In recent years, single-molecule sensitivity achievable by surface-enhanced Raman spectroscopy (SERS) has been widely reported. We use this to investigate supramolecular host–guest chemistry with the macrocyclic host cucurbit[7]uril, on a few-to-single-molecule level. A nanogap geometry, comprising individual gold nanoparticles on a planar gold surface spaced by a single layer of molecules, gives intense SERS signals. Plasmonic coupling between the particle and the surface leads to strongly enhanced optical fields in the gap between them, with single-molecule sensitivity established using a modification of the well-known bianalyte method. Changes in the Raman modes of the host molecule are observed when single guests included inside its cavity internally stretch it. Anisotropic intermolecular interactions with the guest are found which show additional distinct features in the Raman modes of the host molecule.



Single-molecule spectroscopy techniques have made remarkable contributions in understanding molecular chemistry, revealing details about molecule–surface interactions, molecular geometries, and dynamic processes.^{1,2} Surface-enhanced Raman spectroscopy (SERS) with its specific molecular fingerprinting capability is a powerful analytical tool, requiring only simple experimental setups and sample handling. By placing molecules in strong plasmonic fields, the inherently weak Raman scattering cross sections can be dramatically enhanced. This enables detecting substances at the single-molecule level.^{3–6} Until now, single-molecule SERS has been primarily limited to detection and distinction between different chemical species. With the advent of analytical tools at single-molecule sensitivity,² characterization of intermolecular interactions has also drawn interest.^{7–9} It is therefore important to extend the capabilities of SERS to investigate single-molecule chemical interactions.

A powerful plasmonic construct with a strong field confinement can be realized with the “nanoparticle on mirror geometry” (NPoM),^{10–12} where a gold nanoparticle (AuNP) is spaced above a flat Au surface by a single layer of molecules, establishing a plasmonic gap of only a few nanometers. Because the AuNP induces image charges within the metallic surface, plasmonic coupling similar to a nanoparticle dimer is created (Figure 1a,b). This self-assembled system thus creates a robust and reproducible plasmonic sensing platform for SERS with extremely high field localization and is several orders of magnitude below the classical diffraction limit. Due to the

strong confinement of the generated field enhancement, the interrogated volume extends¹³ over only a few cubic nanometers and has shown promise in enabling down to single-molecule resolution.^{14,15}

We choose the macrocyclic host molecule cucurbit[7]uril (CB[7]) as a model system in this study. CB[7] belongs to a family of barrel-shaped molecular hosts, cucurbit[*n*]urils (CB[*n*]), which exhibit unique structural and chemical molecular recognition properties that make them suitable molecular receptors, particularly in aqueous systems.¹⁶ The CB[7] host is able to bind a variety of “guest” molecules such as cyclic and aromatic compounds, which are of interest for applications including environmental pollutant sensing,¹⁷ materials design,¹⁸ and biomedical analyses.¹⁹ Confined by the cavity volume of CB[7], only one guest molecule is typically encapsulated inside CB[7] at a given time (Figure 1c).

It is known from crystallographic studies that CB[*n*] undergoes structural deformations when its cavity is filled by a guest molecule.^{20,21} Theoretical modeling with molecules such as 1 and 2 also predicts pronounced changes in the diameter of CB[7] when they are included inside its cavity as a guest.^{22,23} These structural transformations are detected as shifts in the Raman vibrational frequencies of CB[*n*] as reported using a solution-based gold nanoparticle aggregate

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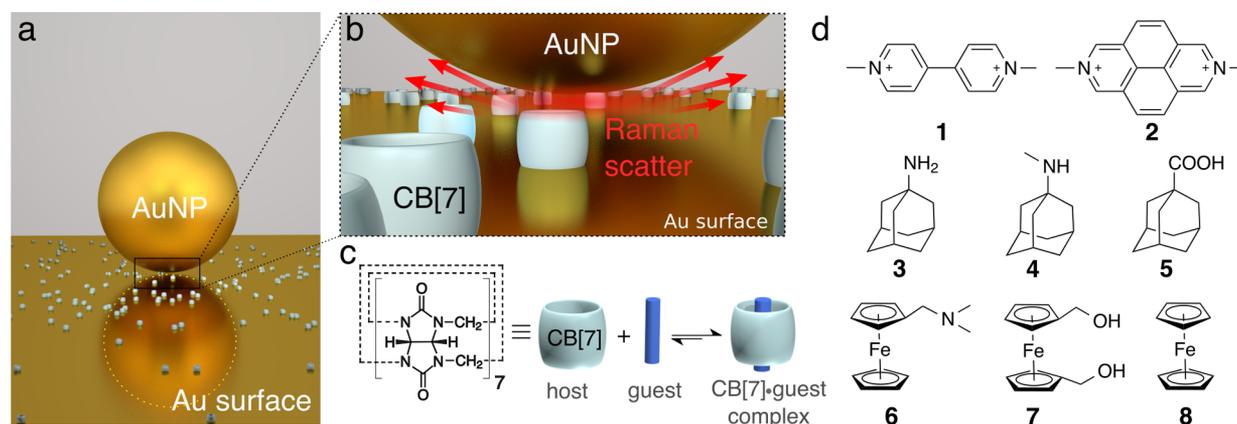


Figure 1. (a) Nanoparticle on mirror assembly. The AuNP is spaced from a gold surface by CB[7] molecules. (b) The plasmonically enhanced field in this construct is confined to a volume of only a few cubic nanometers. (c) The macrocyclic host cucurbit[7]uril accommodates one guest molecule to form a host–guest complex. (d) Guest molecules investigated using the NPoM geometry: 1, methyl viologen; 2, 2,7 dimethyldiazapyrenium; 3, 1-adamantylamine; 4, *N*-methyl-1-adamantylamine; 5, 1-adamantane carboxylic acid; 6, (dimethylaminomethyl)ferrocene; 7, 1,1 ferrocene dimethano; and 8, ferrocene. Note: The counterion for the guests in all cases is chloride.

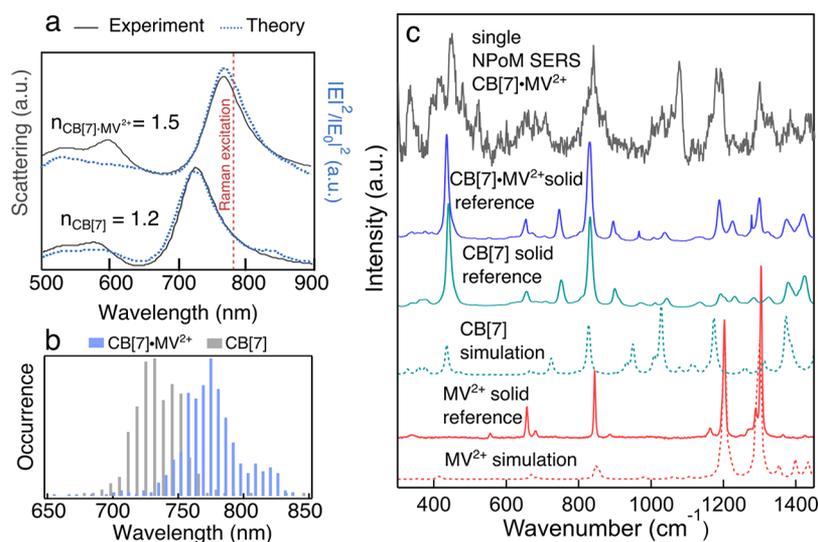


Figure 2. (a) Dark-field scattering spectra of NPoM with MV²⁺·CB[7] and unfilled CB[7] as a spacer. The refractive index change caused by the guest leads to a shift of the plasmon resonance by 30 nm. Dotted (blue) lines are the corresponding field enhancements obtained by modeling for both cases. Vertical (red) dashed line indicates the Raman excitation wavelength ($\lambda = 785$ nm). (b) Statistical distribution of the dark-field scattering resonant wavelengths for unfilled CB[7] (gray) and MV²⁺·CB[7] (blue) as spacers in the NPoM geometry. (c) From top to bottom: MV²⁺·CB[7] SERS spectra on single NPoM, MV²⁺·CB[7] complexed solid-state Raman (experiment), CB[7] solid-state Raman (experiment and simulation), and MV²⁺ solid-state (experiment and simulation).

system.²⁴ It is also possible to study changes in the Raman signatures of guest molecules in bulk (solid) complexes.²⁵

CB[7] is known to adsorb spontaneously on gold surfaces through interactions with the carbonyl portals, giving orientation of its barrel axis perpendicular to the gold surface.²⁶ Here, we use sparsely distributed gold nanoparticles on a planar gold surface separated by a fractional monolayer of complexed CB[7] molecules to fabricate a NPoM geometry with a fixed gap distance of ~ 0.9 nm,²⁷ defined by the barrel height of the CB[7]. Guest molecules 1–8 (see Figure 1d) with reported binding affinities for CB[7] ranging from 10^5 to 10^{16} M⁻¹ were investigated in this study²⁸ (see Supporting Information, section S2). Detailed examination of the Raman modes of CB[7] and its complexes in the gap between particle and surface allows statistical analysis of the host–guest system.

Planar gold surfaces were prepared with a single physisorbed layer of CB[7] of fractional coverage (see Experimental

Section). Colloidal AuNPs (diameter 100 nm) were then drop-cast on the surfaces to form NPoMs with strong field enhancements in the gaps. A distinction between the filled and unfilled states of the CB[7] cavities present within the NPoM plasmonic gaps can readily be observed using dark-field scattering microscopy. For example, the presence of MV²⁺ (1) within the CB[7] cavities shows clear red-shifts in the plasmonic resonances of the NPoM system (Figure 2a,b). The dominant plasmonic mode is a result of the coupling between the AuNP and the surface. These modes are observed at ~ 730 nm when uncomplexed CB[7] is present in the NPoM gap, whereas this mode shifts to ~ 770 nm for gaps containing CB[7]·MV²⁺ instead. This is a result of the higher refractive index of the complexed CB[7].²⁹ The two cases were modeled using the boundary-element method (BEM) with refractive indices of the surrounding medium of $n = 1.2$ for the uncomplexed and $n = 1.5$ for the complexed state to

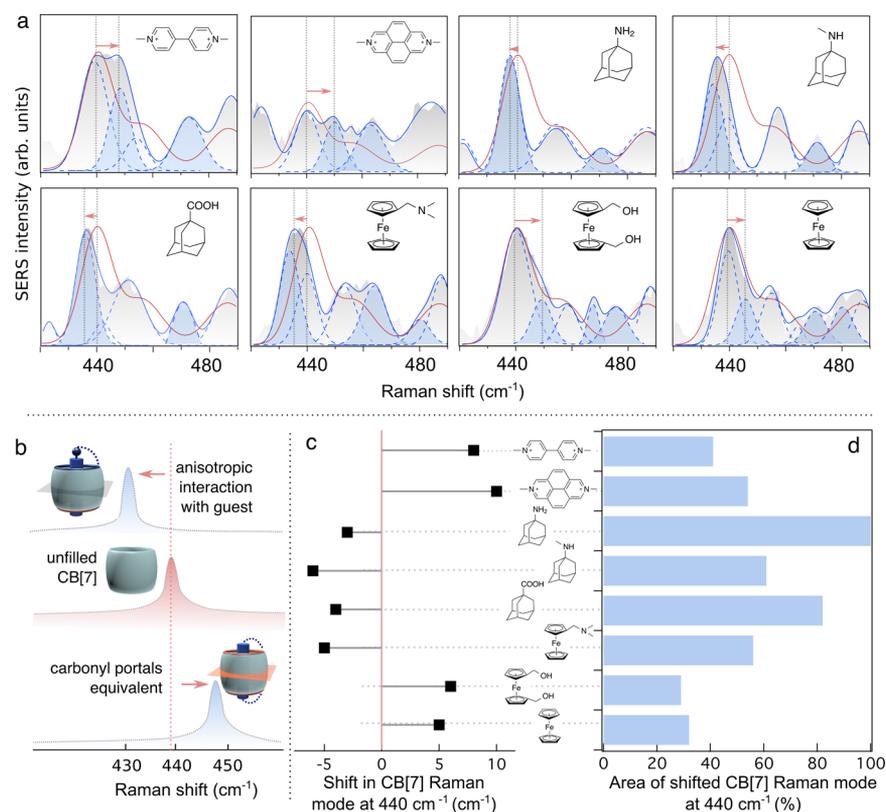


Figure 3. (a) Averaged SERS spectra of the CB[7] Raman modes near 440 cm^{-1} and showing the deconvolution (deconvoluted peaks shown with blue dashed lines), with peaks attributed to filled CB[7] colored blue. The fitted spectra for CB[7] control in the absence of guest has been overlaid (red) in each case for comparison. (b) Schematic showing characteristic trends in shifts. (c) Size of shifts and (d) areas of the shifted CB[7] vibrational mode at 440 cm^{-1} for each guest molecule (% relative to unfilled CB[7]).

qualitatively match the experimental data. The Raman excitation wavelength of 785 nm is thus close to the plasmonic gap resonances, allowing the vibrational spectral positions in the resulting SERS spectra to be compared.

CB[7] has many vibrational modes in the fingerprint region between 1100 and 1500 cm^{-1} with two additionally pronounced peaks that stand out at lower frequencies (Figure 2c). The strongest mode at $\sim 440\text{ cm}^{-1}$ is a “ring scissor mode”, while the other strong mode, observed near 830 cm^{-1} , is attributed to a “ring breathing oscillation”.³⁰ When the macrocycle has a guest encapsulated inside its cavity, these CB[n] breathing modes undergo distortions^{24,25} and provide a sensitive marker to distinguish between the filled and unfilled states of CB[7]. A typical SERS spectrum obtained from a single NPoM (Figure 2c) contains signatures from both CB[7] and MV^{2+} . The Raman bands observed using the NPoM are in agreement with those seen in solid reference samples for both the host and guest molecules as well as with simulations (using HF/3-21G; see Supporting Information, section S1).

Few-molecule spectroscopy is highly sensitive to the immediate environment of the molecules being probed, which leads to inhomogeneous broadening and subtle fluctuations in the peak positions in individual spectra ($\pm 5\text{ cm}^{-1}$). Therefore, many individual spectra (on the order of 300 per molecule) as well as their averaged spectra are inspected. While Raman signals are often weak in individual spectra, they emerge as clearly visible bands in a stacked representation (Supporting Information, Figure S5).

When the Raman spectra of solid CB[7] and that of CB[7] within the NPoM geometry are compared, a new shoulder at

455 cm^{-1} becomes evident. This weak band is consistently observed across all the samples, including for the smaller homologue CB[5], and is likely to originate as a result of CB[n]-Au interactions (see Supporting Information, section S3.2). Aside from this, shifts in the peak positions for the filled and unfilled states of CB[7] show differences depending on the guest molecule encapsulated inside the CB[7] cavity (Figure 3a). An extra signal at $\sim 445\text{ cm}^{-1}$ is also observed for 1, 2, 7, and 8. This signal is, however, notably stronger for planar molecules 1 and 2, which cause a pronounced ellipsoidal deformation of the CB[7] molecule. A different behavior is seen in cases with guests 3–8, where the breathing mode at 440 cm^{-1} is shifted to a lower wavenumber at $\sim 435\text{ cm}^{-1}$. The different shifts to higher or lower wavenumbers arise from the anisotropic interactions of the guest molecule with CB[7], depending on whether the guest molecules interact with only one or both of the two carbonyl portal rims (Figure 3b). A summary of the shifts in the peak position of CB[7] at 440 cm^{-1} for all the studied guests, and the fractional areas of the shifted signals, are shown in Figure 3c,d. In general, the shifted mode is weaker (less area) than the unshifted CB[7] resonance. Guest molecules that interact with both portals exhibit shifts of the 440 cm^{-1} mode to high energy, while those that interact with only one portal shift to lower energy.

A similar trend is not observed for the mode at 830 cm^{-1} because this vibrational mode predominantly arises only from the equatorial region of the CB[7] and therefore is relatively less affected by binding around the carbonyl portals. A slight shift toward lower frequency ($\sim 827\text{ cm}^{-1}$) is observed in all cases, while an additional broad signal can be observed at ~ 870

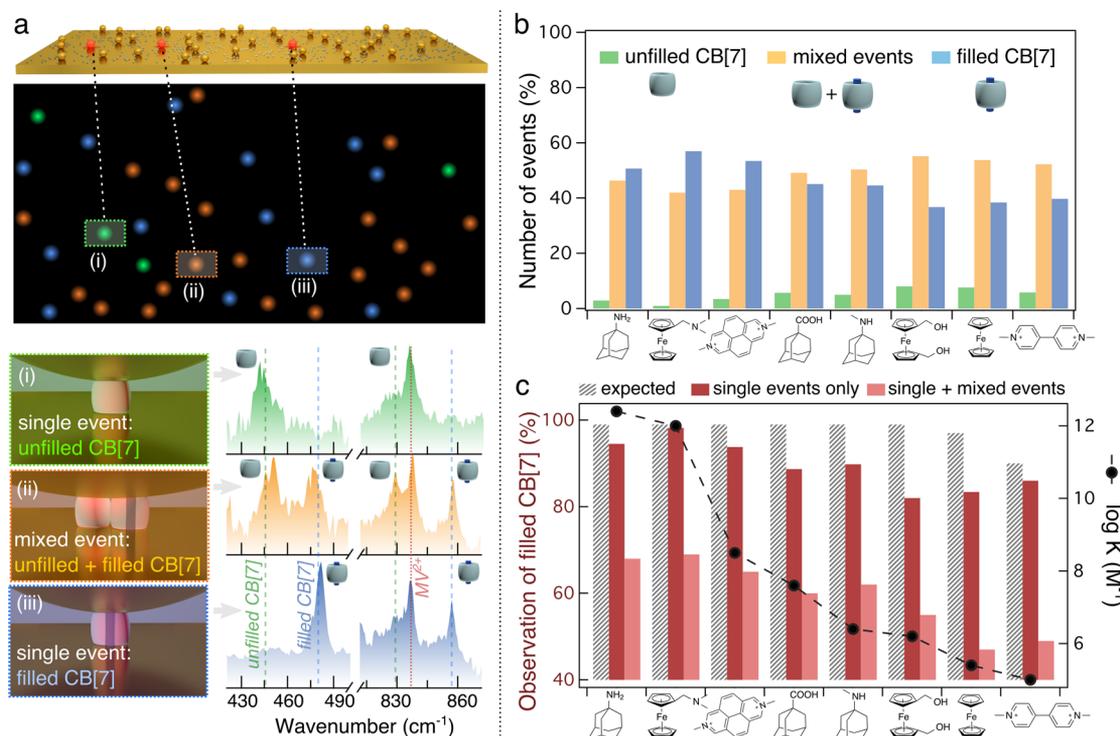


Figure 4. (a) (top) Schematic illustration of distribution of the underlying composition of the CB[7] and MV²⁺·CB[7] mixture in the NPoM probes showing three observable events. (bottom) Individual SERS spectra of the two predominant CB[7] Raman modes near 440 and 830 cm⁻¹ (CB[7] modes are highlighted with gray dashed lines and MV²⁺ modes with red dotted lines). Spectra correspond to (i) NPoM with purely unfilled CB[7] (green), (ii) a mix of filled and unfilled CB[7] (orange), and (iii) only filled CB[7] (blue). (b) Statistical representation of the number of events observed. For each guest molecule, the statistics is based on the investigation of between 300–400 NPoMs with sufficiently strong SERS signals to enable identification of the relevant modes. (c) Comparison of the number of times signals from filled CB[7] is observed for single-molecule events only and mixed events as a function of decreasing order of binding affinities of guests for CB[7]. Overlaid trace shows log of binding strengths of the guest molecules for CB[7].

cm⁻¹ including control samples (of CB[7] and CB[5]). These signals are therefore again likely to arise from the electrostatic interaction of CB[*n*] with the gold surface. While similar magnitudes for such shifts have been shown in SERS using colloidal gold nanoparticle aggregates,²⁴ the different effects of different molecules have not been reported before.

When probing the distribution of the complexed and uncomplexed states of CB[7] in individual NPoMs with only a few molecules in each plasmonic nanogap, three different situations can be observed (Figure 4a): (i) the probed CB[7] molecules are unfilled, resulting in *unshifted* CB[7] Raman modes; (ii) both filled and unfilled states of CB[7] or “mixed events” are detected simultaneously in cases where more than one molecule may be present in the gap; and (iii) all CB[7] molecules in the junction each contain a guest, leading to only the *shifted* Raman modes. Representative spectra showing the 400 and 800 cm⁻¹ regions for **1** are shown in Figure 4b (see Supporting Information, Figures S8–S10 for individual spectra of other guest molecules).

For all the guest CB[7] inclusion complexes, 40%–60% of the investigated NPoMs exhibit either only filled or only unfilled CBs, which strongly suggests that the SERS signal is in many instances generated by single molecules (Figure 4). The mixed events, however, may arise in three cases: (1) More than one molecule is probed. (2) CB[7] host–guest complexes, in general, are known to be dynamic because of the “in-and-out” diffusion of guests from the CB[7] cavity. These diffusional movements are much faster compared to the integration time (10 s) used for measurement of SERS spectra. For example, the

lifetime of MV²⁺·CB[7] is reported to be 5.3 ± 0.5 milliseconds.³¹ Therefore, it is possible that in some cases, depending on the local environment of the complexed molecule, a *temporal averaging effect* is observed due to this diffusion. In these cases, signals may be obtained from the same CB[7] molecule in a filled and unfilled state. (3) Similarly, it is also possible that the incident laser occasionally removes the guest molecules from the CB[7] cavity, also resulting in a temporal average signal being observed. In cases 2 and 3, the mixed event signals could arguably also be attributed to single molecules.

The high number of single-molecule events suggests somehow additional field localization addresses only a few molecules. Possibly this arises from imperfections and adatoms raised above the plain surface in the nanogap architecture that can localize light to even smaller volumes than the gap plasmon and thus play a significant role in promoting selected molecules to produce more intense SERS signals by creating strong locally concentrated electric fields.³²

The method presented here is thus built upon the well-established bianalyte technique, which is used to demonstrate single-molecule sensitivity.^{33–35} Typically, the SERS signal of a mixture of two different chemical species is investigated where the occurrence of only one of the two species in a spectrum is an indication that it originates from only one molecule. In this study, the SERS signals from only the host molecule are monitored, with the *filled* and *unfilled* states of CB[7] (with a guest) now treated as two distinct species. The signatures of the two states are compared via the energy shift of Raman bands

upon complexation. In addition to its simplicity, one of the main advantages of this method is that it enables every active site to be probed and the determination of single-molecule sensitivity without relying on accurate determination of the number of molecules in the nanogap.

Based on the known 1:1 binding stoichiometry and association constants of the guests toward CB[7]^{36–39} (see Table S1), the fraction of CB[7] that should be in a complexed state can be estimated using the Hill equation.⁴⁰ For the concentration used in these experiments, the fraction of CB[7] expected to be in a filled state ranges from ~90% for the weaker binding guests such as **1** and **2** to ~99% for the strong binders (**3–5**). These theoretical fractions were compared with the number of experimental observations of Raman signals from filled CB[7] in approximately 350 spectra (i.e., number of spectra containing shifted CB[7] signals). In general, guests that bind to CB[7] with a strong affinity showed a higher frequency of observations from filled CB[7]. This phenomenon is also evident in the averaged spectra, where these molecules show a weaker band from unfilled CB[7] (Figure 3). Furthermore, a control study using 1:10 MV²⁺:CB[7] (10 μ M MV²⁺:100 μ M CB[7]) in the NPoM geometry showed a significant increase in the number spectra containing empty CB[7] single events compared to the number of filled CB[7] events (Supporting Information, Figure S7). The number of mixed events observed was also lower. These results indicate that the populations of filled and unfilled CB[7] on the gold surfaces are likely not a random distribution of the two states but rather are determined by the binding affinity of the guest toward CB[7]. This is not surprising; the formation of a complex between a host and a guest molecule is a fundamental process in supramolecular chemistry and a variety of interactions and energies are collectively involved.^{40,41} Therefore, the molecules would be expected to reside inside the CB[7] cavity governed by their respective association constants. This observation is consistent with a study investigating the recognition properties of self-assembled monolayers of CB[7] with atomic force microscopy.⁴²

From within the hundreds of spectra analyzed for each guest, only spectra showing either *only filled* CB[7] or *only unfilled* CB[7] (i.e., single events) were selected. More than 80% of these single event spectra were found to originate from *only filled* CB[7] (Figure 4c), which is close to the aforementioned expected range of 90–99%. However, if the *single-event-only* spectra were not preselected and all the collected spectra including mixed events were taken into account, the fraction of filled CB[7] was found to be lower, i.e., 45–60%. (Figure 4c). In our analyses, a mixed event is assumed to originate from *one* filled CB[7] molecule and *one* unfilled CB[7] molecule, whereas the actual number of molecules contributing to the signals in mixed events is unknown. Therefore, the discrepancies in the expected and observed number of filled CB[7] events can be reasonably well accounted for. The probable removal of guest from the CB[7] cavity by the laser light (as suggested earlier) might also lead to the fraction of filled CB[7] being lower than expected.

The intensities of the SERS signals obtained from different NPoMs vary by roughly a factor of 10, despite the well-defined orientation of both molecules and optical fields. A major contributing factor to these variations in SERS intensities is the total number of molecules present in the enhanced field region in the gap, which depends on the precise volume of the cavity formed by the AuNP and substrate, set by the faceting of the

NP. The 100 nm diameter AuNPs used in these experiments exhibit typical facet diameters between ~5 and 50 nm, resulting in up to 100-fold variations of the number of incorporated molecules (see Supporting Information, section S6). Assuming a tightly packed monolayer of CB[7], a small AuNP facet with a 5 nm diameter can accommodate approximately 40 CB[7] molecules inside the nanogap, while a large facet of 50 nm can hold up to 4000 molecules. In this case, however, using dilute solutions of CB[7] to form sparse monolayers, it is estimated that there are typically between 1 and 100 CB[7] molecules in each NPoM gap.

While perfectly spherical AuNPs would lead to similar detection volumes in each NPoM, the actual cavity size varies as a consequence of the pronounced AuNP faceting. Driven by van der Waals interactions, faceted nanoparticles predominantly tend to align their facets parallel to the underlying substrate⁴³ (Supporting Information, Figure S11). This forms a nanoscopic plasmonic cavity,⁴⁴ which supports standing waves with nodes of high and low field enhancements and of sizes defined by the width of the facet. The specific location of the molecules within the gap as well as the in- and out-coupling efficiency of light therefore becomes an important factor in determining the scattering intensity.

Furthermore, the field enhancement and field distribution within the nanogap is highly sensitive to the exact morphology of the NPoM construct.⁴⁵ Inherent nanometer-scale defects exist on the surface of the AuNP as well as on the underlying gold mirror. This creates extremely strong and highly localized fields in particular areas within the gap through an additional lightning-rod effect.⁴⁶ Molecules in the vicinity of these “subhot spots” potentially produce a much higher SERS intensity. Additional intensity variations are also partly caused by alignment mismatches between the laser and the symmetry axis of the NPoM, resulting in fluctuations of the field enhancement.

In conclusion, we show that SERS of individual NPoMs can clearly distinguish between the different kinetic states (filled or unfilled) of the CB[7] molecules within the NPoM plasmonic nanogap. While it is common to use resonant dye molecules with strong Raman responses to demonstrate single-molecule resonant SERS (SERRS),⁴⁷ the results presented here confirm that few-to-single-molecule sensitivity can also be achieved with electronically *nonresonant* molecules.⁴⁸ However, single-molecule resolution is not comprehensively achieved in all NPoMs and it is anticipated that the actual number of traced molecules depends on the exact morphology of the AuNP and the Au surface.

We observe subtle differences in SERS peak positions and their respective broadening or fluctuations depending on the guest bound into the CB[7]. It is also clear that any substituents covalently interacting with only one of the carbonyl portals of CB[7] exerts a considerable influence on its structure. We evidence clear effects from the symmetry of the guest molecules, which should now guide the development of more accurate theoretical models of the host–guest binding configuration. Furthermore, the applicability of this system in direct statistical estimation of the number of filled CB[7] on a surface is demonstrated and may be extended to interrogate host–guest systems in a similar manner. This concept opens up opportunities to establish host–guest chemistry as a single-molecule sensing platform useful for gas sensing, drug detection,⁴⁹ or quantitative sensing of various hazardous substances. Furthermore, it could be used to investigate guest

binding properties for a variety of biologically significant molecules such as peptides, DNA, and lipid membranes and allows real-time observation of chemical reaction kinetics.

EXPERIMENTAL SECTION

Flat gold surfaces were prepared using electron-beam evaporation. A 5 nm thick chromium adhesion layer was deposited on a silicon wafer, followed by a 70 nm thick gold film. Host–guest complex solutions were prepared by mixing guest molecules and CB[7] (100 μ M, 1:1 stoichiometry) followed by sonication for approximately 5 min. Less water-soluble guest molecules (such as ferrocene) required longer sonication times (\sim 30 min). Gold surfaces were functionalized by immersion in the host–guest solution for at least 24 h to allow formation of a distorted and sparsely distributed CB[7] monolayer on the surface. Samples were then rinsed with water and blow-dried. The 100 nm citrate-capped colloidal AuNPs (BBI solutions) were drop cast on the sample surface where physisorption takes place. After 5 min, excess NPs were washed away.

Raman microscopy was performed on a Renishaw inVia Raman microscope using a 100 \times objective with numerical aperture NA = 0.85. To match the plasmon resonance, an excitation wavelength of 785 nm was chosen with optical power in the objective set to 13 mW. Integration time was 10 s. The SERS spectra were collected on single mirror-coupled AuNPs. This was done by identifying individual AuNPs using dedicated bright- and dark-field imaging in order to move them to the center of the objective to which the Raman laser was aligned as well.

SERS signals could not be observed when the Raman laser was not centered on a nanoparticle, i.e., nothing but background noise was observed. Furthermore, there were cases when SERS signals were not obtained even with “on-particle” measurements. This is expected to happen if the nanoparticle being studied is not located on adsorbed molecules as they are only sparsely distributed on the surface. Samples for reference Raman spectra of MV²⁺·CB[7] was obtained by freeze-drying a 1 mM (1:1) solution of the complex.

Optical scattering spectra were taken on an Olympus BX51 research microscope in dark-field configuration using a 100 \times objective with numerical aperture NA = 0.85. The samples were illuminated using a halogen white light source, and scattered light was collected with a multimode fiber in confocal configuration attached to a cooled spectrometer.

The near-field enhancement inside the nanogap was modeled with the boundary element method using BEMAX.^{50,51} The CB[7] and MV²⁺·CB[7] molecular spacers were modeled as a layer of constant refractive index with $n = 1.2$ and $n = 1.5$, respectively,²⁹ between a flat Au surface and a AuNP. The dielectric functions of Au were taken from Johnson and Christy.⁵² The illumination source was a broadband plane-wave, incident at an angle of 58° to account for NA = 0.85 in the experimental setup.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.jpcllett.5b02535.

ITC characterization of CB[7] binding with guest molecules 4 and 7, control studies, additional represen-

tations of the SERS spectra, SEM characterization of sample, and estimation of number of molecules in NPoM gaps (PDF)

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Notes

The authors declare no competing financial interest.

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